### SU-585

# Evaluation of Precision and Accuracy of Nichols Advantage<sup>®</sup> 25-Hydroxyvitamin D Assay for 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub>: Comparison of Four Methods with LC-Mass Spectrometry.\*

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## Abstract

The serum assay for 25-Hydroxyvitamin D [25(OH)D] is used to determine the vitamin D status of patients. The serum 25(OH)D assay measures both 25(OH)D2 and 25(OH)D3. The automated Nichols Advantage® 25(OH)D assay is based on chemiluminescence detection and vitamin D binding protein (DBP) for competitive displacement. To determine how efficient and accurate the Nichols Advantage chemiluminescence assay was for detecting both 25(OH)D2 and 25(OH)D3, as compared to other methods, serum samples were obtained from healthy adults and from patients who were treated for several months with pharmacologic doses of vitamin D<sub>2</sub>. The serum was divided into aliquots and numbered. Frozen samples were provided in a blinded fashion for analysis by four methods: (1) high performance liquid chromatography (HPLC), (2) manual RIA (3) D-binding protein isotopic assay and (4) the Nichols Advantage System. The results were compared to that from liquid chromatography-mass spectrometry (LC-MS/MS). These samples were tested at independent laboratories. A comparison of all the assays suggests that there are differences among the methods. The Nichols Advantage assay was able to detect and quantify 25(OH)D levels in samples containing 25(OH)D, and 25(OH)D,

## Introduction

25-Hydroxyvitamin D measurement is useful in determining the status of vitamin D levels. Deficiency of vitamin D directly affects bone health and a variety of other cellular functions. Vitamin D deficiency is treated either with the natural vitamin D<sub>3</sub> or the synthetic vitamin  $D_3$ . Both are converted to the active 1,25-Dihydroxyvitamin D via first to 25(OH)D. Successful treatment with vitamin  $D_2$  or vitamin  $D_3$  can be determined by the increase in the levels of 25(OH)D. Therefore methods used for determining 25(OH)D need to detect both 25(OH)D2 and 25(OH)D2.

## LC-MS/MS system



## Objective

Compare the 25(OH)D results of four different methods with that of LC-MS/MS on patient samples before and after treatment with pharmacologic doses of Vitamin D<sub>2</sub>.

## Methods for 25(OH)D

#### LC-MS/MS

Sample preparation: For each sample, three 50 microliter aliquots were spiked with 20 ng of internal standard [<sup>2</sup>H]<sub>2</sub>-25(OH)D<sub>3</sub>, allowed to incubate at 37°C for 30 mins and then run through a Cohesive Technologies turbulent flow system followed by laminar flow chromatograghy. Analysis: Detection is by TSQ Quantum Ultra mass spectrometer (Thermo Finnigan Corp.). MS is focused on the transition for  $25(OH)D_3$  and  $25(OH)D_2$  The efficiency of assay for each analyte was corrected by use of an internal standard, [2H],-25(OH)D<sub>a</sub>. Concentration and calibration curves were generated by LC-Quan (Thermo Electron Corp.). Calibration: Calibration curve was generated by analyzing the eight calibration standards in five replicates. Internal Standard: [<sup>2</sup>H]<sub>2</sub>-25(OH)D<sub>3</sub> was synthesized in house from pure 25(OH)D<sub>3</sub> (Calbiochem) with purity >92%.

#### HPLC

Determination of 25(OH)D2 and 25(OH)D3. First, plasma or serum spiked with 8000 cpm [<sup>3</sup>H]-25(OH)D3 was extracted with 2 volumes of acetonitrile, followed by C-18 cartridge chromatography (Hollis BW, Clin Chem 32, 2060-2063, 1986), and finally, normal phase chromatography to separate 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub>

## Manual Radioimmunoassay (RIA)

A commercially available antibody based assay using radio-label and performed by manual extraction procedure (Diasorin)

#### Manual D-Binding Protein Competitive Isotopic Assav

Protein Binding assay was performed according to the meth described previously (Chen TC, Turner AK, Holick MF. J N Biochem, 1:315-319, 1990). Serum or plasma was first extract with 100% ethanol, followed by a protein-binding assay.

#### Nichols Advantage<sup>®</sup> Automated Chemiluminescence Assay

The assay is based on human D-binding protein, anti-human DI labeled with acridinium ester and 25(OH)vitamin D3 bound magnetic particles. The 25(OH)D is released from the bindi protein by a releasing agent and the competitive assay is complet automatically by the Nichols Advantage System.

\* Results for the different methods were obtained from several laboratori

## **Subjects**

Inclusion criteria: Adults 18 or older, able and willing to give informed consent and comply with study protocol.

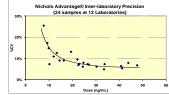
Exclusion criteria: People who are taking medications that can affect vitamin D metabolism.

Treatment: Patients who were vitamin D deficient were treated with 50,000 IU vitaminD<sub>2</sub> once a week for 8 weeks followed by 50,000 IU vitamin D, once every 2-4 weeks for a minimum of 8 weeks.

Samples: Serum obtained from blood samples were made into several aliquots and were frozen for shipment and testing by different methods.

## Inter-assay precision

#### Nichols Advantage®



Nichols Advantage®

Total LC-MS/MS

Range

Nichols Advantage

vs. LC-MS/MS

94%

68%

86%

52%

91%

59%

48%

58%

50%

67 + 18%

48 to 94%

## Results vs. LC-MS/MS in samples predominantly with 25(OH)D<sub>2</sub>

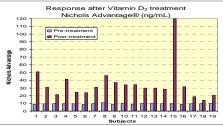
### Manual D-Binding Protein assav

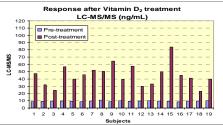
#### D-Binding Protein Assay Total LC-MS/MS D-Binding Protein Assay Nichols Advantage vs. LC-MS/MS (ng/mL) (ng/mL) (na/mL) 53 48 110% 45 40 27 63 158% 43 30 35 106% 30 94% 29 46 49 39 40 98% 19 14 18 24 75% 20 107 + 28% (n=6) Mean + SD 78 to 158%

#### HPI C

hod	TH EV								
Nutr	HPLC	LC-M	S/MS	Total LC-MS/MS	HPLC		RIA	Total LC-MS/MS	
cted	(ng/mL)	25-OH-D <sub>2</sub>	25-OH-D <sub>3</sub>	(ng/mL)	vs. LC-MS/MS		(ng/mL)	(ng/mL)	
	62	37	11	48	129%		59	48	
DBP	43	34	6	40	108%	- E	48	40	_
	47	47	3	50	94%		85	50	
	87	53	5	58	150%		71	58	
	28	24	9	33	85%		37	33	
	47	42	7	49	96%		56	49	
to	38	36	4	40	95%		47	40	
ding	18	20	4	24	75%		29	24	
eted	37	40	0	40	93%		39	40	
	(n=9)			Mean + SD	103 + 23%	_	(n=9)	Mean + SD	
ries.				Range	75 to 150%			Range	

### Response after treatment with Vitamin D<sub>2</sub> [50.000 IU] in vitamin D deficient patients [25(OH)D<10ng/ml]





## Conclusions

- HPLC and the manual D-binding protein methods compared 1 favorably with LC-MS/MS.
- 2. The RIA method shows over estimation.
- Based on LC-MS/MS results. Nichols Advantage® assav can 3 detect and quantify 25(OH)D<sub>o</sub>. Some samples show under estimation
- Nichols Advantage® method can determine the increase in serum 25(OH)D levels in vitamin D-deficient patients who were treated with pharmacologic doses of vitamin D<sub>a</sub>.

#### (ng/mL) 48 40 50 58 33 49 40 24 40 Mean + SD

Range

(n=9)	Mean +		
	Range		
	RIA		

- RIA vs. LC-MS/MS 123% 120% 170% 122% 112% 114% 118%
  - 121% 4 98% 122 + 20% 98 to 170%